

iGEM 2016: Team Pittsburgh  
**Week 20 Lab Notebook**

*Monday, October 3*

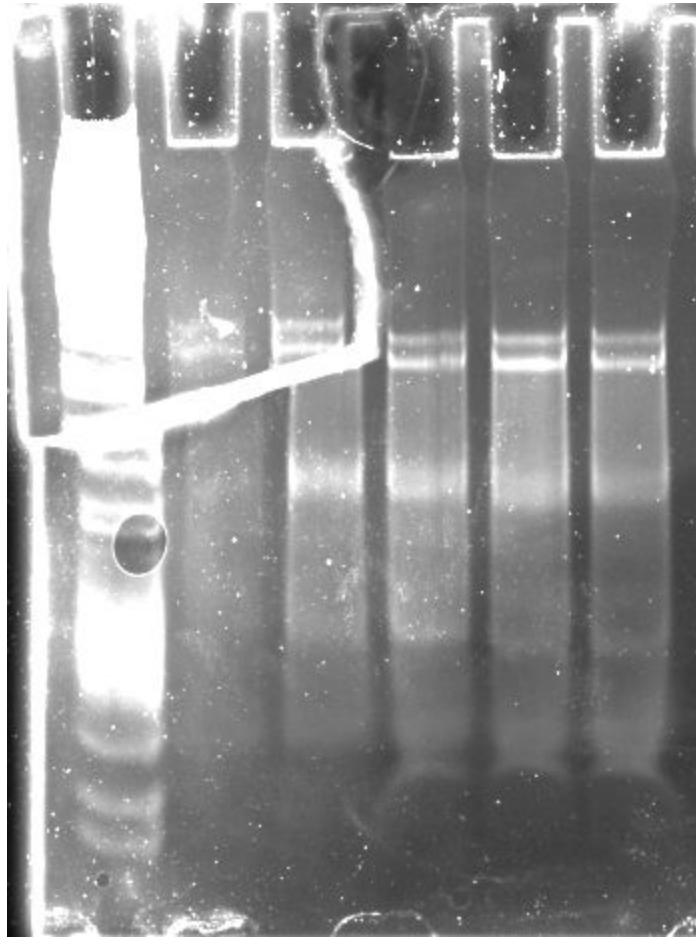
Cleavage reactions (Claire)

50 nM renatured DNAzyme	2.5 uL	1 uM DNAzyme
2 uM lead	2.5 uL	40 uM lead
Buffer (3) or water	<u>45 uL</u>	
	50 uL	

Incubate at 37 for 105 min

*Tuesday, October 4*

PAGE analysis (Praneeth)



*Wednesday, October 5*

Maddie pcr

*Friday, October 7*

Check PT3-GFP and PT7-T3 (Claire)

Digest:

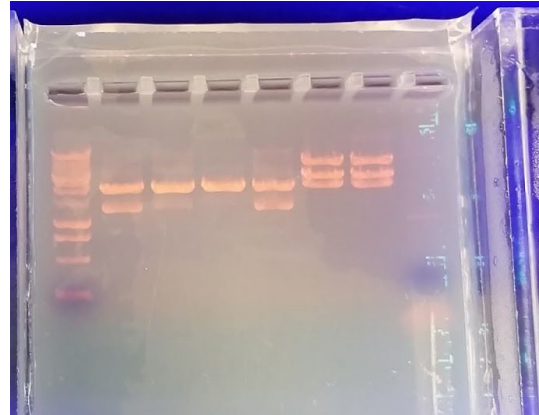
0.5 uL EcoRI  
0.5 uL PstI  
1 uL buffer  
1 ug plasmid  
10 uL with water

DpnI digest of lacZ (Claire)

Incubate 1 hr 30 min at 37

1% agarose gel, 100 V, 37 min (Claire)

Lane	1	ladder
	2	PT7-T3 1
	3	PT7-T3 2
	4	PT7-T3 4
	5	PT7-T3 5
	6	PT3-GFP 3:1
	7	PT3-GFP 7:1
	8	DpnI digested lacZ



Extract lacZ (Claire)

Gel slice didn't fully dissolve. Added another aliquot of extraction buffer to undissolved gel after liquid was transferred to column (ethanol had been added). lacZ 2 is second go

Concentrations: lacZ 1: 0.8 ng/uL  
lacZ 2: -0.3 ng/uL

Ligation (Aife)

Add 2 uL T4 DNA ligase buffer  
7 uL water  
1 uL T4 DNA ligase

Transformation (Aife)

Entire reaction into Cheryl's cells

*Saturday, October 8*

Cell-free lead titration

10/7

Saturday, October 27

Cell-free ~~RNA~~ lead titration & mismatch

25 ng switch, 3.74 nM hp

Control: G, 2  $\mu$ M lead.Conditions: ~~D, 2  $\mu$ M lead~~

G, 50 nM lead

72 nM lead

100 nM

250 nM

500 nM

2  $\mu$ M25  $\mu$ M50  $\mu$ M

no trigger, G

~~50~~ 3.74 nM DNA

hairpin only

11 ~~no rxns~~  
conditions, 22 rxns~~no trigger, D~~~~D + 3.74 nM DNA~~~~D + hairpin only -~~~~D + 3.74 nM D DNA~~

Sol'n A 2.4

B 1.5 1.8

RNase 0.3

substrate 0.3

trigger 0.6

switch 0.31

H<sub>2</sub>O 0.29 } 0.6

30 rxns

x15 rxns

36

72

21

54

4.5

9

4.5

9

1.8

18

9.46

18

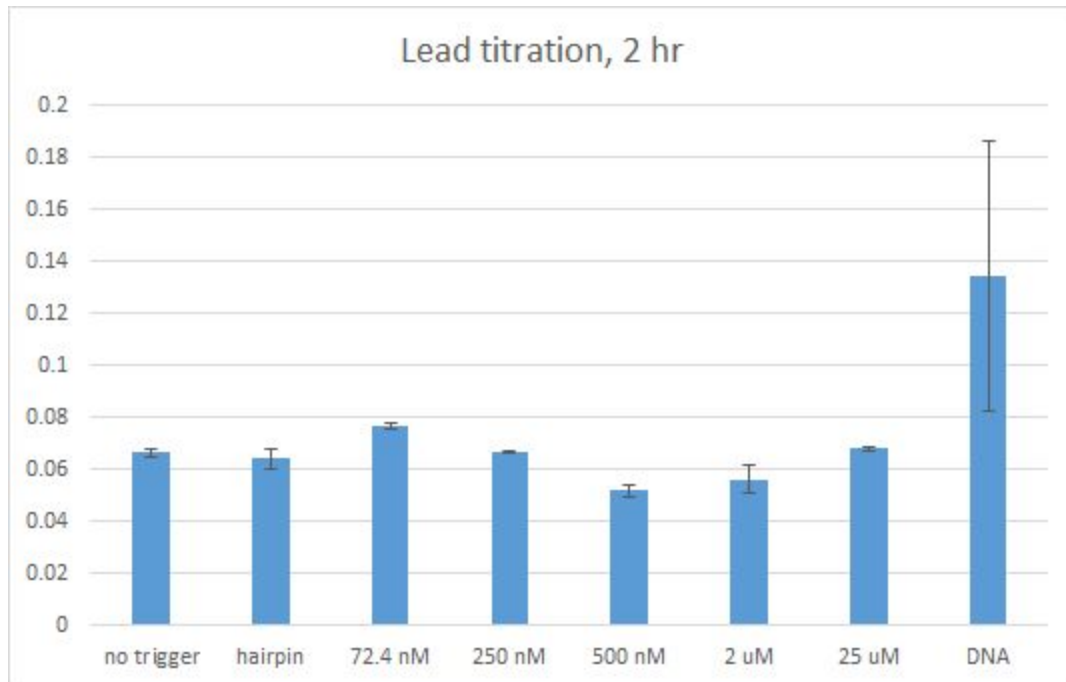
8.54

18

$$\frac{2}{1.448} = 1.38 \quad \begin{matrix} 1 & 10 & 3 \\ ,38 & 3.8 & 1.14 \end{matrix}$$

10/7<sup>111</sup>

TRIGGERS: set up Friday night & freeze		
A1-2	no trigger	1.2 $\mu$ L H <sub>2</sub> O
3-4	hairpin only, 3.74 nM	1 $\mu$ L hp @ 74.8 nM 1 $\mu$ L H <sub>2</sub> O ] 1.2 $\mu$ L hp @ 37.4
<del>3.74 nM hp + 50 nM lead</del>		
<del>1 <math>\mu</math>L hp @ 74.8 nM 1 <math>\mu</math>L Pb<sup>2+</sup> @ 1 <math>\mu</math>M ] 1.2 <math>\mu</math>L</del>		
6 EPA & MCL	3.74 nM hp + 72.4 nM lead	1 $\mu$ L hp @ 74.8 nM 3 $\mu$ L Pb <sup>2+</sup> @ 1.448 $\mu$ M ] 1.2 $\mu$ L
<del>for mismatch experiment (10/10)</del>		
<del>+ 100 nM Pb<sup>2+</sup></del>		
<del>1 <math>\mu</math>L hp @ 74.8 nM 1 <math>\mu</math>L Pb<sup>2+</sup> @ 2 <math>\mu</math>M ] 1.2 <math>\mu</math>L</del>		
7-8	+ 250 nM Pb <sup>2+</sup>	1 $\mu$ L hp @ 74.8 nM 1 $\mu$ L Pb <sup>2+</sup> @ 5 $\mu$ M ] 1.2 $\mu$ L
9-10	+ 500 nM Pb <sup>2+</sup>	1 $\mu$ L hp @ 74.8 nM 1 $\mu$ L Pb <sup>2+</sup> @ 10 $\mu$ M ] 1.2 $\mu$ L
11-12	+ 2 $\mu$ M Pb <sup>2+</sup>	3 $\mu$ L hp @ 74.8 3 $\mu$ L Pb @ 40 $\mu$ M ] 1.2 $\mu$ L
13-14	+ 25 $\mu$ M Pb <sup>2+</sup>	1 $\mu$ L hp @ 74.8 nM 1 $\mu$ L Pb @ 500 $\mu$ M ] 1.2 $\mu$ L
<del>+ 50 <math>\mu</math>M Pb<sup>2+</sup></del>		
<del>1 <math>\mu</math>L hp @ 74.8 nM 1 <math>\mu</math>L Pb @ 1 mM ] 1.2 <math>\mu</math>L</del>		
15-16	3.74 nM DNA	1.2 $\mu$ L DNA @ 37.4 nM



Everything still yellow--should have let reaction go for a bit longer